

# Changes of plasma insulin, urea, amino acids and rumen metabolites in somatotropin treated dairy cows

F. Cheli<sup>1</sup>, A. Baldi<sup>1</sup>, L. Gregoretti<sup>1</sup>, F. Rosi<sup>2</sup>, D. Cattaneo<sup>1</sup>, and V. Dell'Orto<sup>1</sup>

<sup>1</sup>Institute of Animal Nutrition, <sup>2</sup>Institute of Animal Husbandry, University of Milan, Milan, Italy

Accepted August 29, 1997

**Summary.** An experiment was performed to evaluate the effects of somatotropin on plasma free amino acid, urea and insulin concentrations and rumen fermentation pattern and to assess their relationships. Four Italian Friesian dairy cows fitted with rumen cannulae were used in a switch-back design. Slow releasing recombinant bovine somatotropin (640 mg/cow) was injected every 28 days for two consecutive periods. Rumen fluid and blood samples were collected before and after feeding at 0, 7 and 21 days after rbST injection. Exogenous rbST increased plasma insulin concentration and the insulin response to feeding, and decreased plasma urea and free essential and branched chain amino acid concentrations. rbST did not affect rumen fermentation pattern. No correlation was found between rumen and plasma parameters measured after feeding. Our results are consistent with the notion that the main effect of somatotropin is post-absorptive.

**Keywords:** Amino acids – Somatotropin – Plasma amino acids – Dairy cows

#### Introduction

The galactopoietic effect of both pituitary and recombinant bovine somatotropin (rbST) is well documented (McDowell et al., 1987; Peel and Bauman, 1987). rbST treatment increases milk production in lactating cows, without affecting milk composition, if energy and nitrogen balances of treated animals are positive (Chalupa and Galligan, 1989). Somatotropin increases lipolytic and gluconeogenetic processes, while lipogenesis is decreased (Knapp et al., 1992; Liesman et al., 1995). Current dogma suggest that the galactopoietic effect of rbST is partially mediated by changes in the circulating levels of hormones, such as IGF-1, promoting homeorhetic adaptations and enhancing the availability of energy yielding metabolites and the partition of nutrients to the mammary gland (Sechen et al., 1989; Vernon, 1988). Somatotropin increases the efficiency of nitrogen utilization reducing amino

acid oxidation (Peel and Bauman, 1987). Decreased urinary nitrogen excretion and plasma urea levels have been observed in bST treated dairy cows (Sechen et al., 1989; Hanigan et al., 1992).

Experimental data on rbST influence on rumen fermentation pattern and nutrient digestibility seem to indicate that somatotropin has no effect at the preabsorptive level and support the concept that rbST treated cows have a digestive metabolism similar to untreated cows of similar production (Robinson et al., 1991).

The aim of this study was to evaluate the effects of rbST treatment on plasma free amino acid, urea, insulin concentrations and some ruminal metabolites and to asses their relationships.

# Materials and methods

Cows: experimental design

Four lactating Italian Friesian dairy cows (191 ± 44 days of lactation, 15 kg/d of milk) fitted with rumen cannulae were used. The animals, kept in a tied-stall, were fed at 04.30 a.m. 4kg of concentrate and 3kg of meadow hay and at 09.00 a.m. 15kg of maize silage (31% DM), 3kg of meadow hay and 4kg of concentrate given as total mixed ration (TMR). The chemical composition of the feed ingredients is reported in Table 1.

Recombinant bovine somatotropin in a slow release formulation (640 mg/cow; Somidobove Eli Lilly) was injected at the start of two 28 days consecutive periods. In the first period two animals were treated with rbST and two animals with a saline solution. In the following period the treatments were reversed.

# Sample collection and analyses

Milk production was recorded weekly. Milk samples were collected weekly at consecutive morning and evening milkings and were individually analized for fat (Gerber method) and protein (Kjeldahl method) content. Feed intake was recorded weekly for three consecutive days. For each experimental period, at 0, 7 and 21 days after rbST injection, rumen fluid and blood samples from the jugular vein were collected at 09.00 a.m., before TMR administration, and at 02.00 p.m.

For volatile fatty acid (VFA) analysis, 0.8 ml of 5% (wt/vol) metaphosphoric acid was added to 4 ml of rumen fluid and stored at -20°C. VFA concentrations, using 4-methylvaleric acid as internal standard, were determined by gas chromatography with a 60/80 Carbopack S 0.3% Carbowax 20M/0.1% H<sub>2</sub>PO<sub>4</sub> column.

**Table 1.** Chemical composition of feeds (% of dry matter)

	Maize silage	Meadow hay	Concentrate
Dry matter %	31.00	89.00	87.10
Crude protein	7.42	9.98	18.48
Crude fiber	23.54	28.57	7.46
NDF	51.20	70.12	12.80
Fat	3.20	2.90	4.47
Ash	5.52	8.04	7.46

NDF neutral detergent fiber.

Plasma insulin concentration was determined by a commercially available radio-immunoassay kit (Amersham, Italia, S.r.l.).

To measure plasma amino acid concentration (AA), samples of  $200\mu$ l of plasma, with  $20\mu$ l of norleucine added as an internal standard, were deproteinized with  $800\mu$ l of methanol. After centrifugation,  $200\mu$ l of the supernatant was collected in microfuge tubes dried under a stream of N<sub>2</sub>-gas and resuspended in  $100\mu$ l of Li citrate buffer (pH 2.4). Plasma AA concentrations were determined by HPLC with a LiCl ion exchange column. AA were post-column derivatized with o-phthaldehyde and fluorimetrically detected (Condon, 1986).

# Statistical analysis

Rumen and plasma data were statistically analyzed using a least-squares ANOVA program (SAS, 1988). The statistical model included rbST treatment, day from rbST injection and sampling time as main effects. Individual AA and their combinations into total (TAA), essential (EAA), non-essential (NEAA), branched chain (BCAA), ketogenic (KETO) and glucogenic (GLUCO) amino acids were considered. Pearson's correlation analysis was carried out between rumen and plasma parameters.

### Results

Overall mean daily milk yield was significantly higher in rbST-treated than in control cows (18.3  $\pm$  1.9  $\nu$ . 15.4  $\pm$  1.9 kg/d; P < 0.01). There was no effect of rbST treatment on milk protein and fat content. A peak of milk production was recorded 7 days after rbST injection (data not shown).

There was no effect of rbST on overall mean dry matter intake (17.72  $\pm$  1.16kg/cow -control group-  $\nu$ . 17.76  $\pm$  1.04kg/cow -rbST group-).

rbST treatment did not affect rumen VFA molar concentration and the molar proportion of acetate and propionate (Table 2). Ruminal VFA concentrations were higher, although not statistically significant, five hours after TMR administration both in control ( $105 \pm 5 v$ .  $116 \pm 6 \text{mmol/l}$ ) and treated cows ( $117 \pm 6 v$ .  $124 \pm 6 \text{mmol/l}$ ).

Overall mean plasma insulin concentration was significantly higher (P < 0.01) in rbST-treated than in control cows (Table 3). A significant post-prandial increase (P < 0.01) in plasma insulin concentration was found in both treated and control cows (Figure 1). At seven days after rbST injection, insulin response to feeding was amplified in treated animals (Figure 1). In rbST-treated cows, plasma insulin concentrations tended to be higher than in

**Table 2.** Effect of rbST on rumen parameters (lsmeans  $\pm$  SEM)

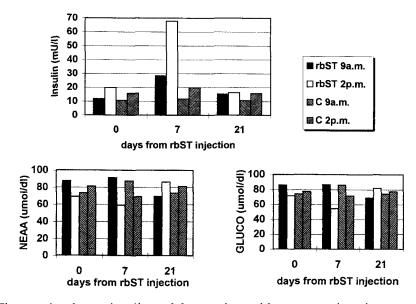
Items	Control	rbST
VFA, mmol/l	$110.42 \pm 3.95$	$120.93 \pm 4.09$
Acetate, % Propionate, %	$58.69 \pm 1.41$ $23.99 \pm 1.34$	$58.20 \pm 1.46$ $25.51 \pm 1.39$

VFA volatile fatty acids.

**Table 3.** Effect of rbST treatment on plasma parameters (Ismeans  $\pm$  SEM)

	Control	rbST	P=
Insulin, mU/L	$14.49 \pm 4.06$	$31.90 \pm 4.20$	0.007
Urea, mmol/l	$2.64 \pm 0.17$	$1.23 \pm 0.09$	0.009
Amino acids, µmol/dl			
Glutamic acid	$3.71 \pm 0.29$	$3.60 \pm 0.28$	0.79
Serine	$6.88 \pm 0.44$	$6.64 \pm 0.43$	0.70
Glycine	$29.13 \pm 1.89$	$27.48 \pm 2.08$	0.56
Glutamine	$21.58 \pm 0.98$	$21.01 \pm 1.02$	0.69
Histidine	$3.53 \pm 0.31$	$2.58 \pm 0.33$	0.05
Threonine	$6.82 \pm 0.52$	$6.14 \pm 0.53$	0.37
Alanine	$12.92 \pm 1.22$	$14.82 \pm 1.35$	0.31
Tyrosine	$4.05 \pm 0.40$	$3.40 \pm 0.42$	0.28
Valine	$19.67 \pm 0.93$	$16.52 \pm 0.96$	0.03
Methionine	$2.05 \pm 0.10$	$2.02 \pm 0.11$	0.84
Isoleucine	$9.50 \pm 0.65$	$7.97 \pm 0.67$	0.12
Leucine	$11.77 \pm 1.00$	$9.76 \pm 1.04$	0.18
TAA	$131.07 \pm 6.00$	$119.01 \pm 6.21$	0.18
EAA	$53.20 \pm 3.12$	$44.69 \pm 3.23$	0.07
NEAA	$77.86 \pm 3.47$	$76.52 \pm 4.04$	0.80
BCAA	$40.96 \pm 2.49$	$34.27 \pm 2.58$	0.07
KETO	$32.16 \pm 2.37$	$27.29 \pm 2.45$	0.17
GLUCO	$77.32 \pm 3.26$	$72.96 \pm 3.80$	0.39

TAA total amino acids; EAA essential amino acids; NEAA non essential amino acids; BCAA branched chain amino acids; KETO ketogenic amino acids; GLUCO glucogenic amino acids.



**Fig. 1.** Changes in plasma insulin and free amino acid concentrations in somatotropin-treated (*rbST*) and control (*C*) cows at 0, 7 and 21 days from rbST injection. 9 a.m., 2 p.m.: sampling time. *NEAA* non essential amino acids; *GLUCO* glucogenic amino acids

the control group at 09.00 a.m.  $(28.4 \pm 8.7 \text{ v.} 11.7 \pm 6.8 \text{mU/ml})$  while at 02.00 p.m., five hours after TMR, the difference was statistically significant  $(67.7 \pm 8.4 \text{ v.} 19.9 \pm 8.7 \text{mU/ml}; P < 0.01)$ . rbST treatment decreased (P < 0.01) plasma urea concentration (Table 3).

Overall means of plasma concentrations of some AA and their combination according to their catabolic fates are reported in Table 3. No significant effect of time of the day on plasma free AA leves was found. rbST treatment decreased plasma concentrations of His, Val (P < 0.05), EAA and BCAA (P = 0.07). This effect was statistically more significant at seven days after rbST injection, when the peak in milk production was recorded, for both EAA (54.79  $\pm$  4.17 $\mu$ mol/dl – control cows-  $\nu$ . 40.07  $\pm$  4.50 $\mu$ mol/dl -rbST-treated cows; P < 0.05) and BCAA (42.24  $\pm$  3.30 $\mu$ mol/dl -control cows-  $\nu$ . 31.30  $\pm$  3.56 $\mu$ mol/dl -rbST-treated cows; P < 0.05). In rbST-treated cows, a significant decrease in NEAA and GLUCO after TMR administration was found at seven days after rbST-injection (Figure 1). A negative correlation between insulin and NEAA was found at 02.00 p.m. (r = -0.71; P < 0.06).

Five hours after TMR administration no correlations were found between rumen and plasma parameters.

# Discussion

In this study, rbST treatment increased milk production with no change in protein and fat content. Changes in milk composition of rbST-treated cows have been attributed to a status of negative energy and protein balance (Chalupa and Galligan, 1989). The weekly response in milk production showed a peak at 7 days after rbST injection in accordance with previous reports (Oldenbroek et al., 1989; Cattaneo et al., 1993).

Dry matter intake did not differ between rbST-treated and control cows. This is consistent with the results of other authors (Peel and Bauman, 1987) who reported that long-term administration of bST gradually increased feed intake, but that there appeared to be 4 to 5 week lag prior to its full expression. A substantial increase in feed consumption was found only after 12 weeks of bST supplementation (Chalupa and Galligan, 1989).

In the present study rbST treatment did not affect rumen fermentation parameters in agreement with previous reports (Dell'Orto and Savoini, 1991; Winsryg et al., 1991). A higher VFA absorption in rbST-treated cows has been described (Savoini et al., 1992). These changes may be related to the greater energy requirements associated with enhanced milk production.

Plasma insulin concentration was significantly higher in rbST-treated than in control cows in accordance with previous reports (McDowell et al., 1987; Wanderkooi et al., 1995), however no effect of somatotropin on insulin concentration has been found by other authors (Cisse et al., 1991). The increased insulin concentration in rbST-treated animals may be due to a compensatory secretion related to a reduced tissue sensitivity to insulin (Walton et al., 1986; Magri et al., 1990). This impairment of insulin action and the consequently reduced glucose uptake by peripheral tissues should facilitate the preferential utilization of glucose by the mammary gland (Ronge and Blum, 1989). The

significant post-prandial increase in insulin concentrations observed in this study is in agreement with other studies and may be induced by feed intake, through the stimulation of the entero-insular axis, and VFA absorption (Morgan et al., 1988; Sutton et al., 1988). The post-prandial changes of insulin concentration were higher in treated than in control cows at seven days after rbST injection, when the peak in milk production is recorded and increased plasma somatotropin concentration is reported (Dell'Orto et al., 1993). Since feed intake and rumen fermentation pattern were not different, we might consider that the regulation of nutrient partitioning by homeorhetic and homeostatic mechanisms may be different in rbST-treated and control animals.

Plasma urea concentration was lower in treated than in control animals. The decreased urea levels in bST-treated cows may be an expression of decreased gluconeogenic use of AA and improved utilization of nitrogen (Peel and Bauman, 1987; Sechen et al., 1989; Hanigan et al., 1992). Plasma free AA concentrations were affected by bST treatment, in accordance with previous studies (Fullerton et al., 1989; Baldi et al., 1990; Hanigan et al., 1992). EAA and BCAA concentrations were lower in treated cows. Treatmentinduced changes were more evident at seven days after rbST injection, when the peak in milk production was recorded. Once absorbed in the blood, AA are immediately subject to metabolism by the liver, which has a profound impact on their availabilty to the mammary gland. The liver metabolizes individual AA to varying degrees and the net liver removal of EAA and BCAA is relatively low (Reynold et al., 1994). The observed reduced plasma concentrations of EAA and BCAA in rbST-treated cows may indicate a higher uptake by the mammary gland to support higher milk protein synthesis. Synthesis and secretion of milk protein within the udder might be promoted by two processes: 1) increased quantity of AA taken up by the mammary gland and 2) improved efficiency of conversion of plasma EAA to milk protein in the secretory cells (Guinard and Rulquin, 1995). How these two factors interact is still not known. Furthermore the effect of bST in reducing plasma AA concentration may be due to an increased blood flow to the mammary gland (Davis et al., 1988). The observed significant decrease of plasma levels of NEAA and GLUCO after TMR administration in bSTtreated cows may be related to the high plasma insulin concentration found.

In conclusion, bST treatment did not affect rumen metabolites, but significantly influenced plasma concentration of insulin, urea and free amino acids. Treatment-induced changes were more evident at seven days after rbST injection, when the peak in milk production was recorded. Observed changes in plasma metabolites induced by rbST appeared to be unrelated to changes in rumen metabolites. These results are consistent with the notion that the main effect of somatotropin is post-absorptive.

# References

Baldi A, Rosi F, Cheli F, Savoini G, Dell'Orto V, Salimei E (1990) bST treatment in dairy cows fed different levels of concentrate: effects on plasma free amino acids. Proceedings of the 41st Annual Meeting European Association Animal Production, Toulose 1990, p 176 (Abstr)

- Cattaneo D, Salimei E, Savoini G, Baldi A, Cheli F, Fantuz F, Dell'Orto V (1993) Effetti dell'impiego di somatotropina bovina e dei saponi di calcio su produzione e composizione del latte. Zoot Nutr Anim 4: 217–228
- Chalupa W, Galligan DT (1989) Nutritional implications of somatotropin for lactating cows. J Dairy Sci 72: 2510–2524
- Cisse M, Chillard Y, Coxam V, Davicco MJ, Remond B (1991) Slow release somatotropin in dairy heifers and cows fed two levels of concentrate. 2. Plasma hormones and metabolites. J Dairy Sci 74: 1382–1394
- Condon GC (1986) Amino acid analysis. Theory and laboratory techniques. LKB Biochrom Limited, Cambridge, England, p 63
- Davis SR, Collier RJ, Mc Namara JP, Head HH, Sussman W (1988) Effects of thyroxine and growth hormone treatment of dairy cows on milk yield, cardiac output and mammary blood flow. J Anim Sci 66: 70–79
- Dell'Orto V, Savoini G (1991) Recombinant bovine somatotropin (rbST) treatment in dairy cows: effect on ruminal activity and milk properties. Microbiologie-Aliments-Nutrition 9: 121–132
- Dell'Orto V, Savoini G, Salimei E, Cattaneo D, Secchi C, Rosi F (1993) Effects of recombinant bovine somatotropin (rbST) on productive and physiological parameters related to dairy cow welfare. Livest Prod Sci 36: 71–75
- Fullerton FM, Mepham TB, Fleet JR, Heap RB (1989) Changes in mammary uptake of essential amino acids in lactating Jersey cows in response to exogenous bovine pituitary somatotropin. In: Heap RP, Prosser CG, Lamming GE (eds) Biotechnology in growth regulation. Butterworths, London, p 239
- Guinard J, Rulquin H (1995) Effects of graded amount of methionine on the mammary uptake of major milk precursors in dairy cows. J Dairy Sci 78: 2196–2207
- Hanigan MD, Calvert CC, DePeters EJ, Reis BL, Baldwin RL (1992) Kinetic of amino acids extraction by lactating mammary glands in control and Sometribove-treated Holstein cows. J Dairy Sci 5: 161–173
- Knapp JR, Freetly HC, Reis BL, Calvert CC, Baldwin RL (1992) Effects of somatotropin and substrates on patterns of liver metabolism in lactating dairy cattle. J Dairy Sci 75: 1025–1036
- Liesman JS, McNamara JP, Capuco AV, Binelli M, Vanderkooi WK, Emery RS, Tucker HA, Moseley WM (1995) Comparison of growth hormone-releasing factor and somatotropin: lipid and glucose metabolism in dairy cows. J Dairy Sci 78: 2159–2166
- Magri KA, Adamo M, LeRoith D, Etherton TD (1990) The inhibition of insulin action and glucose metabolism by porcine growth hormone in porcine adipocites is not the result of any decrease in insulin binding or insulin receptor kinase activity. Biochem J 266: 107–113
- McDowell GH, Hart IC, Bines JA, Lindsay DB, Kirby AC (1987) Effects of pituitary-derived bovine growth hormone on production parameters and biokinetics of key metabolites in lactating dairy cows at peak and mid lactation. Austr J Biol Sci 40: 191–202
- Morgan LM, Flatt PR, Marks V (1988) Nutrient regulation of the entero-insular axis and insulin secretion. Nutr Res Rev 1: 79–98
- Oldenbroek JK, Garssen GJ, Forbes AB, Jonker LJ (1989) The effect of treatment of dairy cows of different breeds with recombinantly derived bovine somatotropin in a sustained-delivery vehicle. Livest Prod Sci 21: 13–34
- Peel CJ, Bauman DE (1987) Somatotropin and lactation. J Dairy Sci 70: 474-486
- Reynold CK, Harmon DL, Cecava MJ (1994) Absorption and delivery of nutrients for milk synthesis by portal-drained viscera. J Dairy Sci 77: 2787–2808
- Robinson PH, DeBoer G, Kennelly JJ (1991) Effect of bovine somatotropin and protein on rumen fermentation and forestomach and whole tract digestion in dairy cows. J Dairy Sci 74: 3505–3517
- Ronge H, Blum JW (1989) Insulin-like growth factor I response to recombinant bovine growth hormone during feed restriction in heifers. Acta Endocr (Copenhagen) 120: 735–744

- Savoini G, Van Vuuren AM, Van Der Koelen CJ (1992) The effects of daily administration of recombinantly-derived somatotropic hormone on feed intake, digestibility, milk composition and rumen fermentation in dairy cows. J Anim Physiol Anim Nutr 68: 185–196
- Sechen SJ, Bauman DE, Tyrrel HF, Reynolds PJ (1989) Effect of somatotropin on kinetics of nonesterified fatty acids and partition of energy, carbon and nitrogen in lactating dairy cows. J Dairy Sci 72: 59–67
- Sutton JD, Hart IC, Moran SV, Schuller E, Simmond AD (1988) Feeding frequency for lactating cows: diurnal pattern of hormones and metabolites in peripheral blood in relation to milk fat concentration. Br J Nutr 60: 265–274
- SAS/STAT (1983) User's Guide Statistics, Ed Cary, NC, SAS Institute., Inc. USA
- Vernon RY (1988) Influence of somatotropin on metabolism. In: Sejersen K, Vestergaard M, Neimann Sorensen A (eds) Use of somatotropin in livestock production. Elsevier Applied Science, London, New York, pp 31–50
- Walton PE, Etherton TD, Evock CM (1986) Antagonism of insulin action in cultured pig adipose tissue by pituitary and recombinant porcine growth hormone: potentiation by hydrocortisone. Endocrinology 118: 2577–2581
- Wanderkooi WK, Vandehaar MJ, Sharma BK, Binelli M, Tucker HA, Akers RM, Moseley WM (1995) Comparison of growth hormone-releasing factor and somatotropin: the somatotropic axis in lactating primiparous cows. J Dairy Sci 78: 2140–2149
- Winsryg MD, Arambel MJ, Kent BA, Walters JL (1991) Effect of sometribove on rumen fermentation, rate of passage, digestibility and milk production responses in dairy cows. J Dairy Sci 74: 3518–3523

**Authors' address:** Dr. Federica Cheli, Istituto di Alimentazione Animale, Facolta' di Medicina Veterinaria, Universita' di Milano, Via Trentacoste 2, I-20134 Milano, Italy.

Received March 20, 1997